Guidelines for the Use of Antiretroviral Agents in Pediatric HIV Infection

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Laboratory monitoring of children living with HIV poses unique and challenging issues. In particular, normal ranges and the value of CD4 T lymphocyte (CD4) cell count and plasma HIV-1 RNA concentration (viral load) for prediction of risk of disease progression vary significantly by age. This section will address immunologic, virologic, general laboratory, and clinical monitoring of children with HIV, relevant to both those who have recently received an HIV diagnosis and those who are receiving antiretroviral therapy (ART).

### Clinical and Laboratory Monitoring of Children Living With HIV

**Initial Evaluation of Newly Diagnosed Children**

Children who have recently received an HIV diagnosis should have their CD4 cell counts and plasma viral loads measured, and their growth and development evaluated for signs of HIV-associated abnormalities. They should also undergo a laboratory evaluation that looks for HIV-associated conditions, including anemia, leukopenia, thrombocytopenia, hypoalbuminemia, nephropathy (urinalysis), and elevated levels of glucose, transaminases, or creatinine. In addition, children with HIV should have a complete age-appropriate medical
history and physical examination (see Table 3). Opportunistic infection (OI) monitoring should follow guidelines appropriate for the child’s exposure history and clinical setting (see the Pediatric Opportunistic Infections Guidelines).

Laboratory confirmation of HIV infection should be obtained if available documentation is incomplete (see Diagnosis of HIV Infection). Genotypic resistance testing should be performed, even if ART is not initiated immediately. In addition, a full antiretroviral (ARV) drug history, including exposure to ARVs for the prevention of perinatal HIV transmission, should be obtained (see Antiretroviral Drug-Resistance Testing in the Adult and Adolescent Antiretroviral Guidelines). If abacavir is being considered as part of the regimen, HLA-B*5701 testing should be sent prior to initiation of that ARV drug, and an alternative ARV drug should be used if HLA-B*5701 testing is positive (see Abacavir in Appendix A: Pediatric Antiretroviral Drug Information).

Readiness for ARV drug adherence should be assessed prior to starting ART and associated discussion/counseling implemented.

If a child is not placed on ART after HIV diagnosis, monitoring of CD4 count and plasma viral load should be implemented at least every 3 to 4 months.

**Evaluation at Initiation of Combination Antiretroviral Therapy**

At the time of ART initiation, CD4 cell count and plasma viral load should be measured to establish a baseline to monitor ART response. To set the baseline for monitoring ART toxicity (see Management of Medication Toxicity or Intolerance), complete blood count (CBC) and differential, serum chemistries (including electrolytes, creatinine, glucose, hepatic transaminases), urinalysis, and serum lipids (cholesterol, triglycerides) should be measured. CBC allows monitoring of zidovudine-associated anemia, leukopenia, and macrocytosis (see Zidovudine in Appendix A: Pediatric Antiretroviral Drug Information). Electrolytes with anion gaps might help identify nucleoside reverse transcriptase inhibitor-associated lactic acidosis. With use of tenofovir disoproxil fumarate (TDF), creatinine may increase, phosphate may decrease, and proteinuria can occur (see Tenofovir Disoproxil Fumarate in Appendix A: Pediatric Antiretroviral Drug Information). Use of protease inhibitors may be associated with hyperglycemia. Hepatic transaminases (alanine aminotransferase and aspartate aminotransferase) increase with many ARV drugs. Bilirubin should be measured prior to starting atazanavir, because that drug causes an increase in indirect bilirubin (see Atazanavir in Appendix A: Pediatric Antiretroviral Drug Information). For further details of adverse effects (AEs) associated with a specific ARV drug, see Tables 15a-15l in Management of Medication Toxicity or Intolerance.

**Clinical and Laboratory Monitoring After Initiation of Combination Antiretroviral Therapy (or After a Change in Antiretroviral Therapy)**

Children who start ART or who change to a new regimen should be followed to assess effectiveness, tolerability, and AEs of the regimen and to evaluate medication adherence. Frequent patient visits and intensive follow-up during the initial months after a new ART regimen is started are necessary to support and educate the family. The first few weeks of ART can be particularly difficult for children and their caregivers; they must adjust their schedules to allow for consistent and routine administration of medication doses. Children may also experience AEs of medications, and both children and their caregivers need assistance to determine whether the effects are temporary and tolerable or are more serious or long-term and require a visit to the clinician. It is critical that providers speak to caregivers and children in a supportive, non-judgmental manner using layman’s terms. This promotes honest reporting and ensures that providers can have a productive dialogue with both children and their caregiver(s), even when medication adherence is reported to be inconsistent.

**Within 1 to 2 Weeks of Initiation of Antiretroviral Therapy**

Within 1 to 2 weeks of initiating therapy, children should be evaluated either in person or by phone to identify clinical AEs and to support adherence. Many clinicians plan additional contacts (in person, by
telephone, or via email) with children and caregivers to support adherence during the first few weeks of therapy.

2 to 4 Weeks after Initiation of Antiretroviral Therapy

Most experts recommend laboratory testing at 2 to 4 weeks (and not more than 8 weeks) after initiation of ART to assess virologic response and laboratory toxicity, though this recommendation is based on limited data. The selection of laboratory chemistry tests is regimen-specific (see above). Plasma viral load monitoring is important as a marker of response to ART because a decline in viral load suggests medication adherence, administration of appropriate doses, and viral drug susceptibility. Some experts favor measuring viral load at 2 weeks to ensure that viral load is declining. A significant decrease in viral load in response to ART should be observed by 4 to 8 weeks of therapy.

Clinical and Laboratory Monitoring for Children Who are Stable on Long-Term Antiretroviral Therapy

After the initial phase of ART initiation, regimen adherence, effectiveness (CD4 cell count and plasma viral load), and toxicities (history, physical, and laboratory testing as above) should be assessed every 3 to 4 months in children receiving ART. Children who develop symptoms of toxicity should have appropriate laboratory evaluations. If laboratory evidence of toxicity is identified, testing should be performed more frequently until the toxicity resolves.

Table 3 provides one proposed general monitoring schedule, which should be adjusted based on the specific ART regimen a child is receiving.

CD4 count improvement is influenced by the baseline value at ART initiation; children with very low CD4 counts may take longer than 1 year to achieve their highest values after viral load suppression. Recent studies have critically evaluated the frequency of laboratory monitoring in both adults and children, particularly CD4 cell count and plasma viral load. These studies support less frequent monitoring in stable patients in whom viral suppression has been sustained for at least 1 year.

The current Adult and Adolescent Guidelines support plasma viral load testing every 6 months for individuals who have both:

- Consistent virus suppression for longer than 2 years
- CD4 count consistently >300 cells/mm^3

The Panel on Antiretroviral Therapy and Medical Management of Children Living with HIV finds value in continuing viral load testing every 3 to 6 months to provide enhanced monitoring of adherence or disease progression among children and youth. Some experts monitor CD4 cell count less frequently (e.g., every 6 to 12 months) in children and adolescents who are adherent to therapy and have CD4 cell count values well above the threshold for OI risk, sustained viral suppression, and stable clinical status for more than 2 to 3 years. Some clinicians find value in visits every 3 months even when lab testing is not performed, in order to review adherence and update dosing for interim growth.

Testing at the Time of Switching Antiretroviral Therapy

When a switch in regimen is made to simplify ART, labs appropriate to the toxicity profile of the new regimen should be measured at baseline. Follow-up should include plasma viral load at 4 weeks (and not more than 8 weeks) after the switch to ensure efficacy of the new regimen. If the regimen is switched because of ART failure (see Recognizing and Managing Antiretroviral Treatment Failure in Management of Children Receiving Antiretroviral Therapy), resistance testing should be performed while a patient is still receiving the failing regimen. This optimizes the chance of identifying resistance mutations because resistant strains may revert to wild type within a few weeks of stopping ARV drugs (see Antiretroviral Drug-
Resistance Testing in the Adult and Adolescent Antiretroviral Guidelines). Among children with prolonged or repeated periods of viral nonsuppression in the face of serial ART regimens, phenotypic resistance testing, including co-receptor tropism testing, should be considered in addition to genotypic viral resistance testing.11

Immunologic Monitoring in Children: General Considerations

Clinicians interpreting CD4 cell count and percentage in children must consider age as a factor. CD4 cell count and percentage values in healthy infants without HIV are considerably higher than values observed in adults without HIV (and slowly decline to adult values by age 5 years). An analysis from the HPPM Collaborative Study found that CD4 percentage provided little or no additional prognostic value compared with CD4 cell count regarding short-term disease progression in children aged <5 years, as well as in older children.12 The current pediatric HIV disease classification is based on absolute CD4 cell count, which is the preferred assay for monitoring and estimating risk for disease progression and OIs.13

In children living with HIV, as in adults living with HIV, the CD4 cell count and percentage decline as HIV infection progresses; patients with lower CD4 cell count/percentage values have a poorer prognosis than patients with higher values (see Tables A–C in Appendix C: Supplemental Information).

While guidelines now recommend that children of all ages and adults receive ART regardless of CD4 count and clinical stage, CD4 count-associated risk profiles contribute to the level of urgency for recommendations on when to initiate therapy in a treatment-naive child with HIV infection and when to assess need for OI prophylaxis (see When to Initiate). A meta-analysis from the HPPM Collaborative Study generated plots which can be used to estimate the short-term risk of progression to AIDS or death in the absence of effective ART according to age and the most recent CD4 percentage/absolute CD4 cell count or HIV-1 RNA viral load measurement.14

Measurement of CD4 cell count and percentage can be associated with considerable intrapatient variation.15 Mild intercurrent illness, the receipt of vaccinations, or exercise can produce a transient decrease in CD4 cell count and percentage; thus, CD4 cell count and percentage are best measured when patients are clinically stable. Clinical decisions, especially those concerning therapy changes, should be made in response to confirmed changes in CD4 cell count/percentage in conjunction with a confirmed viral load determination, with a minimum of 1 week between measurements.

HIV RNA Monitoring in Children: General Considerations

Quantitative HIV-1 RNA assays measure the plasma concentration of HIV RNA as copies/mL, commonly referred to as the plasma viral load. Without therapy, plasma viral load initially rises to high peak levels during the period of primary infection in adults and adolescents, and then it declines by as much as 2 to 3 log_{10} copies to reach a stable lower level (the virologic set point) approximately 6 to 12 months after acute infection.16,17 In adults living with HIV, the stable lower level (or viral set point) correlates with the subsequent risk of disease progression or death in the absence of therapy.18

The pattern of change in plasma viral load in untreated infants with perinatal HIV infection differs from that in adults and adolescents with HIV infection. High plasma viral load persists in untreated children for prolonged periods.19,20 In one prospective study of infants with perinatal infection who were born prior to ARV drug availability for children, plasma viral loads generally were low at birth (i.e., <10,000 copies/mL), increased to high values by age 2 months (most infants had values >100,000 copies/mL, ranging from undetectable to nearly 10 million copies/mL), and then decreased slowly, with a mean plasma viral load of 185,000 copies/mL during the first year of life.21 After the first year of life, plasma viral load slowly declined over the next few years.21-24 Viral load during the first 12 to 24 months after birth showed an average decline of approximately 0.6 log_{10} copies/mL per year, followed by an average decline of 0.3 log_{10} copies/mL per year until age 4 to 5 years. This pattern probably reflects the lower efficiency of an immature but developing immune system in containing viral replication and possibly the rapid expansion of HIV-susceptible cells that occurs with somatic growth.25
Despite data indicating that high plasma viral load is associated with disease progression, the predictive value of specific HIV RNA concentrations for disease progression and death for an individual child is moderate.\textsuperscript{23} Plasma viral load may be difficult to interpret during the first year of life because values are high and are less predictive of disease progression risk than in older children.\textsuperscript{20} In both children and adults living with HIV, CD4 cell count or percentage and plasma viral load are independent predictors of disease progression and mortality risk, and use of the two markers together more accurately defines prognosis.\textsuperscript{23,24,26,27}

\textbf{Methodological Considerations in Interpretation and Comparability of HIV RNA Assays}

Based on accumulated experience with currently available assays, viral suppression is currently defined as a plasma viral load below the detection limit of the assay used (generally <20 to 75 copies/mL). This definition of suppression has been much more thoroughly investigated in adults with HIV than in children with HIV (see the \textit{Adult and Adolescent Antiretroviral Guidelines}).\textsuperscript{28} Temporary viral load elevations (“blips”) between the level of detection and 500 copies/mL often are detected in adults\textsuperscript{29} and children on ART and should not be considered to represent virologic failure as long as the values return to below the level of detection at the time of repeat testing. For definitions and management of virologic treatment failure, see Recognizing and Managing Antiretroviral Treatment Failure in \textit{Management of Children Receiving Antiretroviral Therapy}. These definitions of viral suppression and virologic failure are recommended for clinical use. Research protocols or surveillance programs may use different definitions.

Several different methods can be used for quantitating HIV RNA, each of which has a different level of sensitivity (see \textit{Table 4}). Although the results of the assays are correlated, the absolute HIV RNA copy number obtained from a single specimen tested by two different assays can differ by two-fold (0.3 log\textsubscript{10} copies/mL) or more.\textsuperscript{30,31} Because of the variability among assays in techniques and quantitative HIV RNA measurements, a single HIV RNA assay method should be used consistently to monitor an individual patient when possible.\textsuperscript{32-34}

The predominant HIV-1 subtype in the United States is subtype B—the subtype for which all initial assays were targeted. Current kit configurations for all companies have been designed to detect and quantitate essentially all viral subtypes (see \textit{Diagnosis of HIV Infection}). This is important for many regions of the world where non-B subtypes are predominant, as well as for the United States, where a small subset of individuals contract non-B viral subtypes.\textsuperscript{32,35-39} It is particularly relevant for children who are born outside the United States or to foreign-born parents.

Biologic variation in plasma viral load within one person is well documented. In adults, repeated measurement of plasma viral load using the same assay can vary by as much as three-fold (0.5 log\textsubscript{10} copies/mL) in either direction over the course of a day or on different days.\textsuperscript{26,31} This biologic variation may be greater in infants and young children with HIV. This inherent biologic variability must be considered when interpreting changes in plasma viral load in children. Thus, on repeated testing, only differences greater than five-fold (0.7 log\textsubscript{10} copies/mL) in infants younger than 2 years and greater than three-fold (0.5 log\textsubscript{10} copies/mL) in children aged 2 years and older should be considered reflective of plasma viral load changes that are biologically and clinically significant.

Generally, no change in ARV treatment should be made as a result of a change in plasma viral load unless the change is confirmed by a second measurement. Interpretation of plasma viral load for clinical decision making should be done by or in consultation with an expert in pediatric HIV infection because of the complexities of HIV RNA testing and the age-related changes in plasma viral load in children.

\textbf{Genetic Testing for Management of HIV}

The evaluation of human and pathogen genes is increasingly being employed to manage disease intervention, and this approach to treatment is featured in the rise of precision medicine. Clinicians who manage HIV have routinely probed HIV’s genetic sequences for mutations associated with HIV drug resistance. Some ARV drugs are metabolized differently based on specific human genotypes. For example, studies have shown
that certain genotypes can affect efavirenz exposure in young children. In addition, some human genetic polymorphisms are associated with drug toxicity or adverse events (e.g., using HLA-B*5701 testing to predict abacavir hypersensitivity). Future clinical practice is likely to feature broader applications of multiple forms of genetic testing to guide management of health and disease.

Table 3. Sample Schedule for Clinical and Laboratory Monitoring of Children Before and After Initiation of Antiretroviral Therapy

<table>
<thead>
<tr>
<th></th>
<th>Entry Into Care&lt;sup&gt;a&lt;/sup&gt;</th>
<th>Pre-Therapy&lt;sup&gt;b&lt;/sup&gt;</th>
<th>ART Initiation&lt;sup&gt;c&lt;/sup&gt;</th>
<th>Weeks 1–2 on Therapy</th>
<th>Weeks 2–4 on Therapy</th>
<th>Every 3–4 Months&lt;sup&gt;d&lt;/sup&gt;</th>
<th>Only Required Every 6–12 Months&lt;sup&gt;e&lt;/sup&gt;</th>
<th>ARV Switch</th>
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<td>Plasma Viral Load</td>
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<td>Chemistries&lt;sup&gt;f&lt;/sup&gt;</td>
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<tr>
<td>Random Plasma Glucose&lt;sup&gt;g&lt;/sup&gt;</td>
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<td>✓</td>
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<tr>
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</tr>
</tbody>
</table>

<sup>a</sup> See text for details on recommended laboratory tests to obtain.

<sup>b</sup> Readiness for ARV adherence is assessed prior to starting ART. If abacavir is being considered as part of the regimen, send HLA-B*5701 testing prior to initiation of that ARV and choose an alternative ARV if HLA-B*5701 is positive (see Abacavir in Appendix A: Pediatric Antiretroviral Drug Information). Genotype resistance testing is recommended if not already performed (see Antiretroviral Drug-Resistance Testing in the Adult and Adolescent Antiretroviral Guidelines). Send tests appropriate to the toxicities expected from each patient's ART regimen and history (see text).

<sup>c</sup> If ART is initiated within 30 to 90 days of a pre-therapy lab result, repeat testing may not be necessary.

<sup>d</sup> CD4 cell count, CBC, and chemistries can be monitored less frequently (every 6–12 months) in children and youth who are adherent to therapy and have CD4 cell values well above the threshold for opportunistic infection risk, sustained viral suppression, and stable clinical status for more than 2 to 3 years. Viral load testing every 3 to 4 months is generally recommended to monitor ARV adherence.

<sup>e</sup> If lipids have been abnormal in the past, more frequent monitoring might be needed. For patients treated with TDF, more frequent urinalysis should be considered.

<sup>f</sup> Chemistries refer to a comprehensive metabolic panel.

<sup>g</sup> Random plasma glucose collected in a gray top tube.

<sup>h</sup> Recommended when considering starting ARV drugs with activity against hepatitis B, specifically lamivudine-, emtricitabine-, and tenofovir-containing regimens.

<sup>i</sup> Recommended only when individual previously demonstrated no immunity to hepatitis B.

**Key to Acronyms:** ART = antiretroviral therapy; ARV = antiretroviral; CBC = complete blood count; CD4 = CD4 T lymphocyte; TDF = tenofovir disoproxil fumarate
Table 4. Primary, FDA-Approved Assays to Monitor Viral Load

<table>
<thead>
<tr>
<th>Assay</th>
<th>Abbott Real Time</th>
<th>NucliSens EasyQ v 2.0</th>
<th>COBAS Ampliprep/TaqMan v 2.0</th>
<th>Versant v 1.0</th>
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<tbody>
<tr>
<td>Method</td>
<td>Real-time RT-PCR</td>
<td>Real-time NASBA</td>
<td>Real-time RT-PCR</td>
<td>Real-time RT-PCR</td>
</tr>
<tr>
<td>Dynamic Range (copies/mL)</td>
<td>40–10³</td>
<td>25–10³</td>
<td>20–10³</td>
<td>37–11x10³</td>
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<tr>
<td>Specimen volume*</td>
<td>0.2–1 mL</td>
<td>0.1–1 mL</td>
<td>1 mL</td>
<td>0.5 mL</td>
</tr>
<tr>
<td>Manufacturer</td>
<td>Abbott</td>
<td>bioMerieux</td>
<td>Roche</td>
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</tr>
</tbody>
</table>

* Smaller volumes for children can be accommodated.

Key to Acronyms: FDA = Food and Drug Administration; NASBA = nucleic acid sequence-based amplification; RT-PCR = reverse transcription polymerase chain reaction

References


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